Integrating multiple transcript assemblies for improved gene structure annotation with Mikado

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**Decoding Living Systems** 



#### Outline

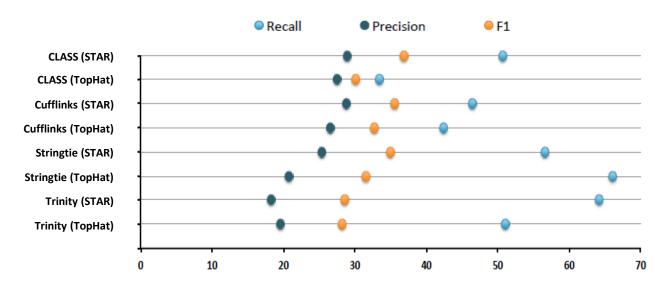
- The first step in many gene builds is to assemble RNA-Seq data to identify reliable gene models to guide *ab initio* predictors or integrator pipelines
- The RGASP challenge in 2013 demonstrated that transcript reconstruction is still an open challenge, both for *ab initio* predictors and RNA-Seq assemblers
- In this talk we will:
  - Show how RNA-Seq assemblers exhibit complementary strengths and weaknesses, making hard to choose *a priori* the best method
  - Present a novel method to integrate multiple transcript assemblies into a clean evidence-based gene build with Mikado





## Varying accuracy of assemblers and aligners

Following the RGASP challenge, we decided to assess the efficiency of different combinations of aligners and assemblers on the three species analysed on the challenge (*C. elegans*, *D. melanogaster* and *H. sapiens*) and additionally the model plant organism *A. thaliana*.



Recall / Precision / F1 %

Recall = proportion of transcripts correctly called

Precision = proportion of called transcripts that are correct

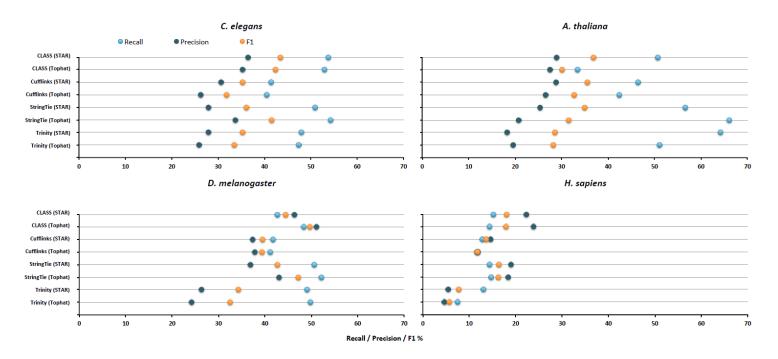
F1 = harmonic mean of recall and precision





## Varying accuracy of assemblers and aligners

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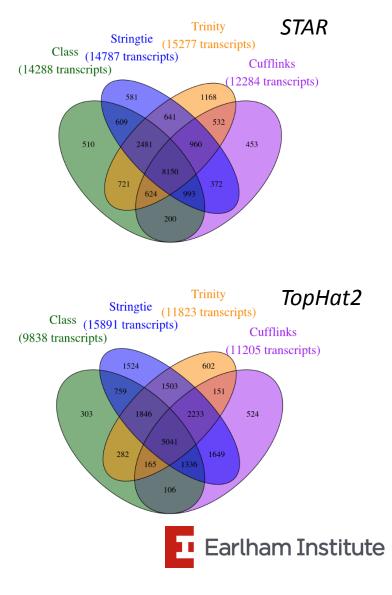
# Complementarity of different assemblers

Fully reconstructed transcripts in *A. thaliana* (from 22,577 transcripts with min 1X coverage and all splice junctions identified)

Method	Fully reconstructed transcripts	% of transcripts reconstructed
Stringtie (Tophat)	15,891	70.39%
Trinity (STAR)	15,277	67.67%
Stringtie (STAR)	14,787	65.50%
CLASS (STAR)	14,288	63.29%
Cufflinks (STAR)	12,284	54.41%
Trinity (Tophat)	11,823	52.37%
Cufflinks (Tophat)	11,205	49.63%
CLASS (Tophat)	9838	43.58%
All methods	19,855	87.94%
STAR methods	18,995	79.83%
Tophat methods	18,024	84.13%

3885 transcripts, or <u>17.21%</u>, reconstructed across all 8 methods.

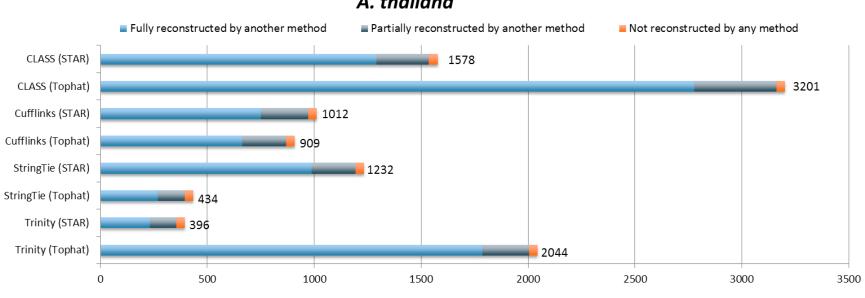
1771 transcripts, or <u>7.85%</u>, reconstructed by only 1 of the 8 methods.





#### **Complementarity of different assemblers: missing genes**

A mirroring problem is that some tools might completely fail to reconstruct the transcript at a locus, even when the available evidence is sufficient for another tool to infer the original transcript(s), or at least part thereof.



A. thaliana

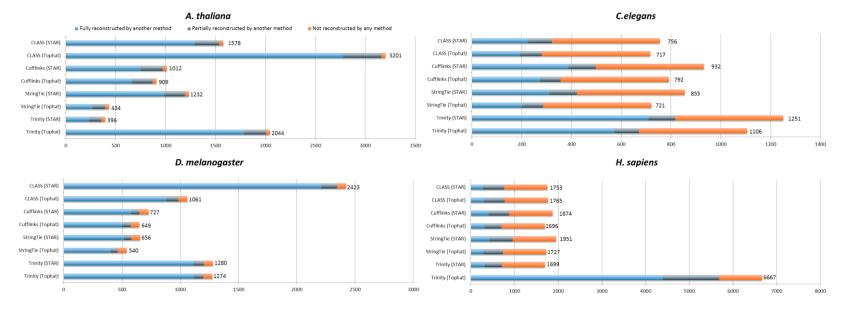
Number of genes





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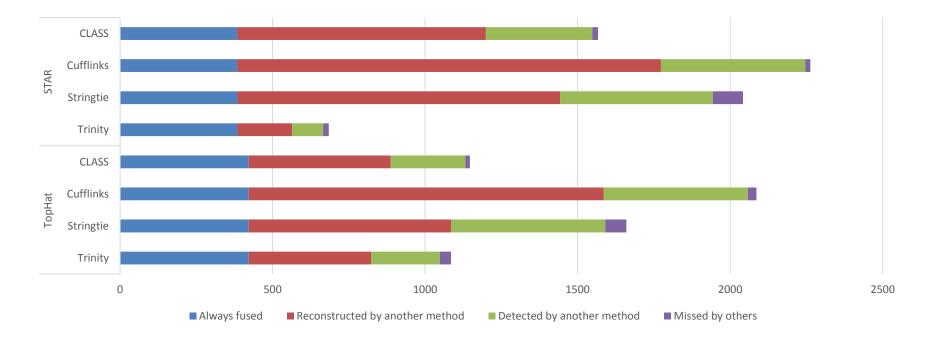
Number of genes





## Artifactual fused genes

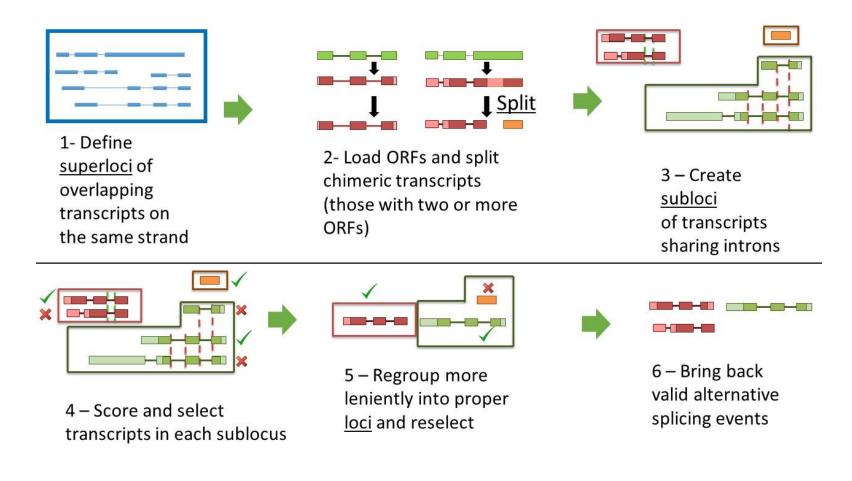
An under-appreciated problem in RNA-Seq assemblies is the tendency to merge together neighbouring genes, especially in compact genomes. Very sensitive methods such as Stringtie can be very prone to this kind of error.







# Leveraging multiple transcript assemblies



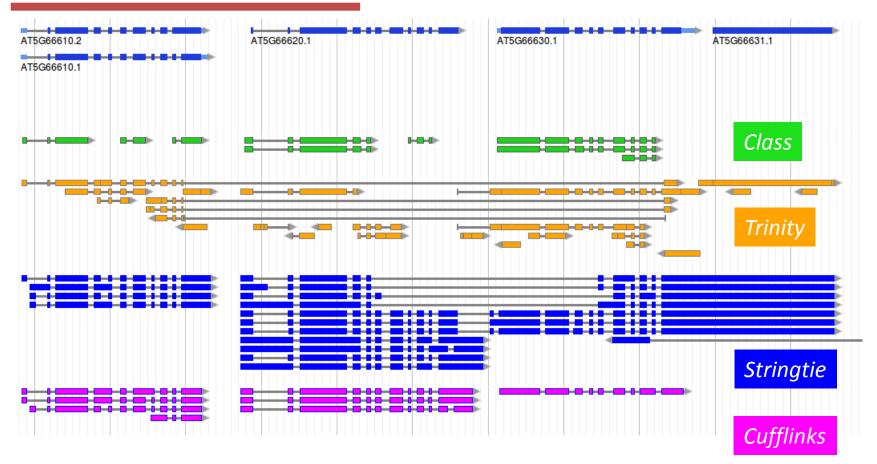




AT5G66610.2	AT5G66620.1	AT5G66630.1	AT5G66631.1
AT5G66610.1			

















# The scoring algorithm

Mikado measures each transcript along over 50 dimensions. The user can specify for each what the program should look for; a final score is computed from the sum of all metrics.

For each metric, the user can decide whether to:

• <u>Maximize</u> its value

$$s_{mt} = w_m st ig(rac{r_{mt} - min(r_m)}{max(r_m) - min(r_m)}ig)$$

• <u>Minimize</u> its value

 $s_{mt} = w_m st \left(1 - rac{r_{mt} - min(r_m)}{max(r_m) - min(r_m)}
ight)$ 

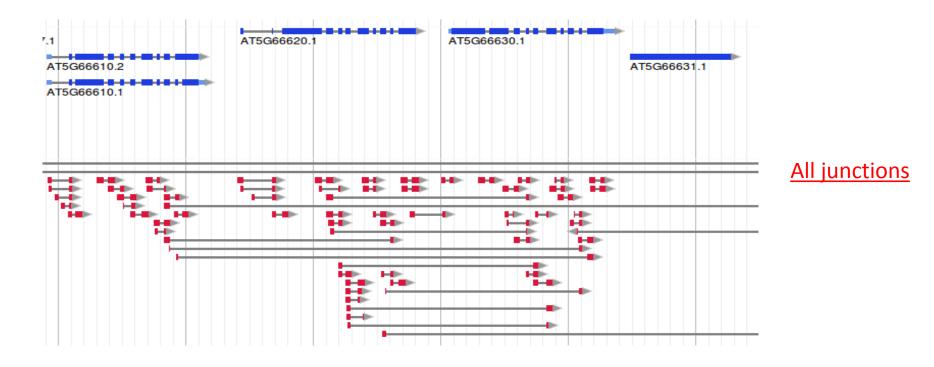
• Look for transcripts with values most similar to a <u>target:</u>

$$s_{mt} = w_m * (1 - rac{|r_{mt} - v_m|}{max(|r_m - v_m|)})$$

Category	Description	Count
External	Data confirmed by external programs, eg. Portcullis	7
Intron	Features related to the number of introns and their lengths	5
cDNA	basic features of any transcript such as its number of exons and its cDNA length	2
CDS	Features related to the ORF(s) assigned to the transcript	24
Locus	features of the transcript in relationship to all other transcripts in its current locus	6
UTR	features related to the UTR of the transcript	12

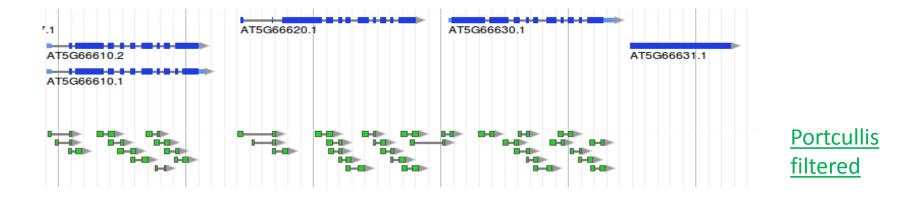








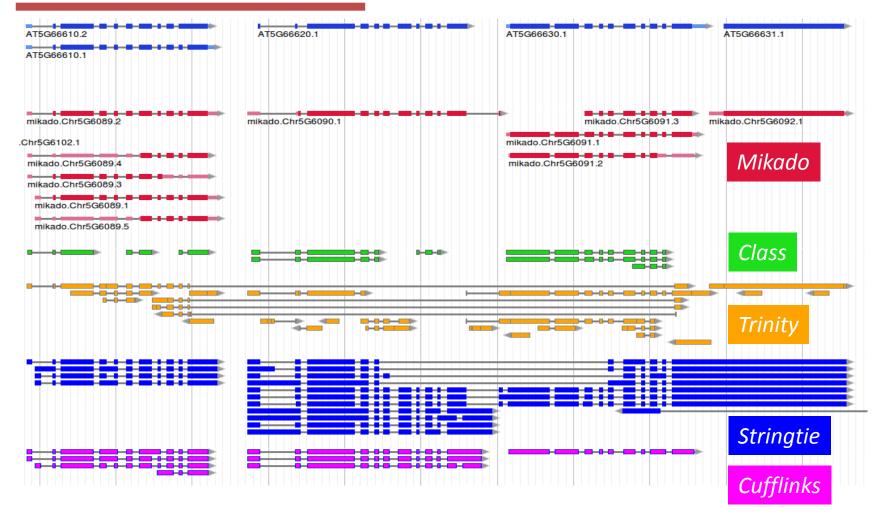








# Solving complex regions with Mikado



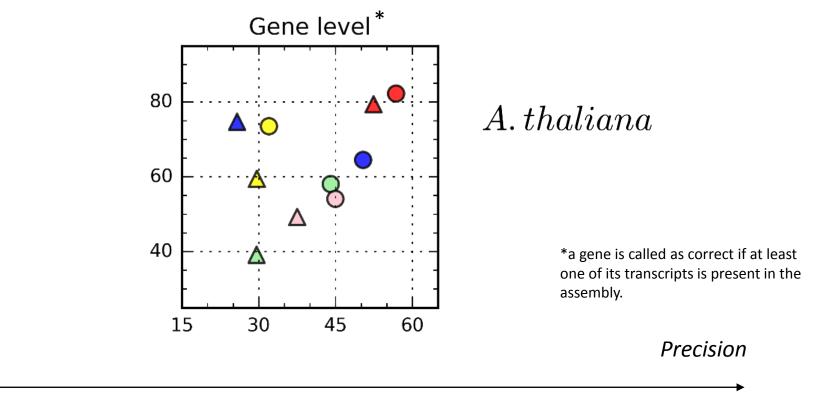




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## Increased precision after filtering with Mikado

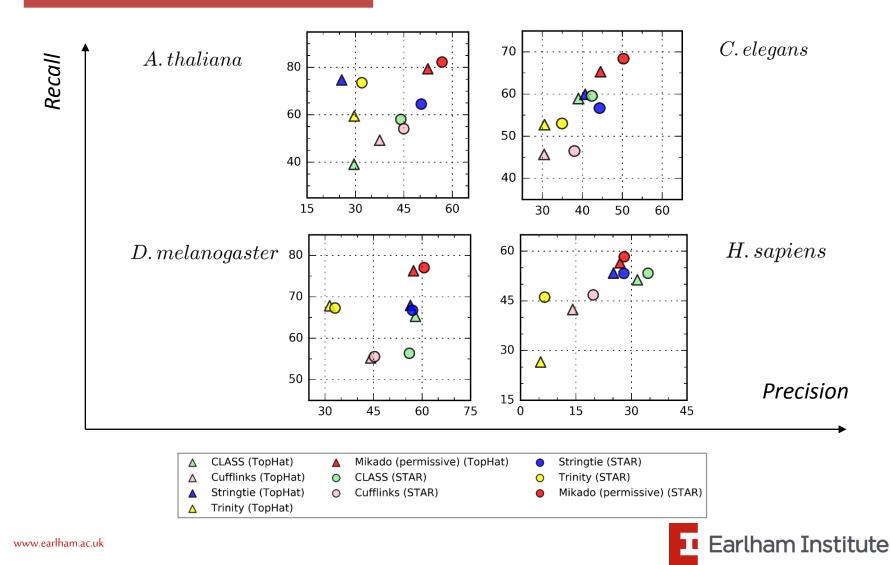
Recall



	CLASS (TopHat) Cufflinks (TopHat) Stringtie (TopHat) Trinity (TopHat)		Mikado (permissive) (TopHat) CLASS (STAR) Cufflinks (STAR)	• •	Stringtie (STAR) Trinity (STAR) Mikado (permissive) (STAR)
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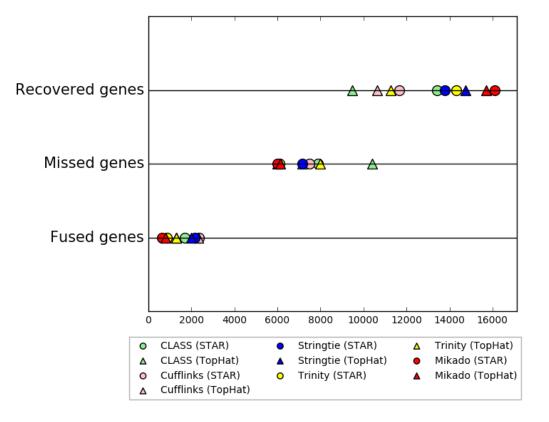


## Increased precision after filtering with Mikado





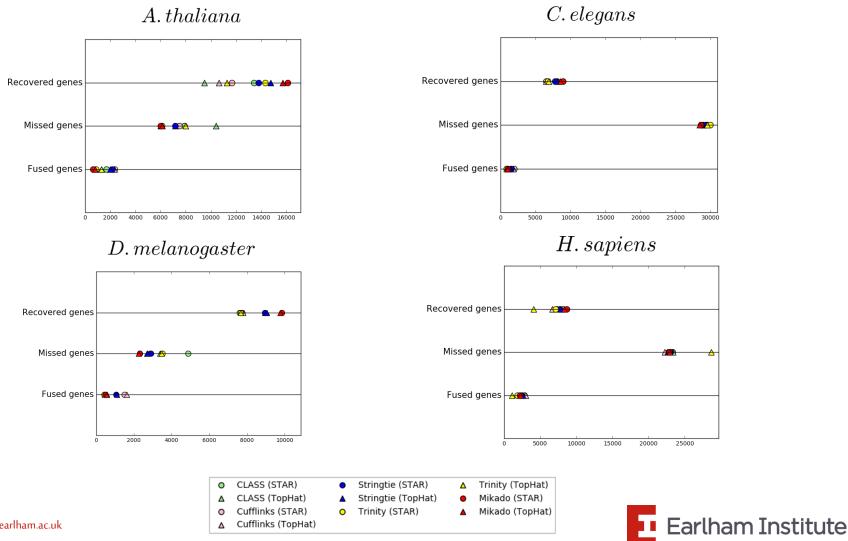
#### Mikado greatly reduces the incidence of fusions in the data







#### Mikado greatly reduces the incidence of fusions in the data





#### Summary

- Mikado is capable of recovering a majority of the annotated isoforms in pooled assemblies.
- The method proposed is completely transparent to the user in terms of what will be used to define acceptable transcripts.
- The software is relatively lightweight and fast the picking operation for *A.thaliana* can be executed in ~15 minutes using approximately 3GB of RAM.
- Mikado uses standard configuration files (JSON/YAML) and file formats (GTF/GFF3/BED):
  - Extendable to other assemblers with standard output formats
- For more information, come visit me at my poster or just go to:



https://github.com/lucventurini/mikado/



https://mikado.readthedocs.org/





#### Acknowledgements



Shabhonam Caim is the primary tester of the pipeline and his assistance has been essential in performing all the experiments presented here



**Gemy George Kaithakottil** verified that the pipeline was effective also on different, nonmodel organisms



**Daniel Mapleson** is the creator of Portcullis and has helped with polishing and improving the efficiency of the pipeline.



**David Swarbreck** is the creator of the pipeline and has shepherded the team throughout the whole development cycle.













# Selecting for general accuracy ...

	C. elegans		A. thaliana		D. melanogaster		H. Sapiens		All species	
	z-score	Rank	z-score	Rank	z-score	Rank	z-score	Rank	z-score	Rank
Method										
CLASS (STAR)	7.30	1	7.61	1	-3.31	6	5.25	1	16.85	1
StringTie (Tophat)	5.51	3	0.59		6.63	2	3.20	3	15.93	2
CLASS (Tophat)	6.98	2	-5.56	<i>XIIIIIIII.</i>	9.33	1	5.00	2	15.75	3
StringTie (STAR)	-2.20	4	2.63		1.59	3	3.00	4	5.01	. 4
Cufflinks (STAR)	-2.32	5	2.70	2	-1.76	5	1.04	5	-0.34	5
Cufflinks (Tophat)	-5.36	,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,	-0.52	5	-1.56	4	-0.99	6	-8.44	6
Trinity (STAR)	-5.07	7	-4.19	7	-4.74	7	-3.42	7	-17.41	
Trinity (Tophat)	-4.83	6	-3.26	6	-6.18	11111111	-13.08		-27.34	





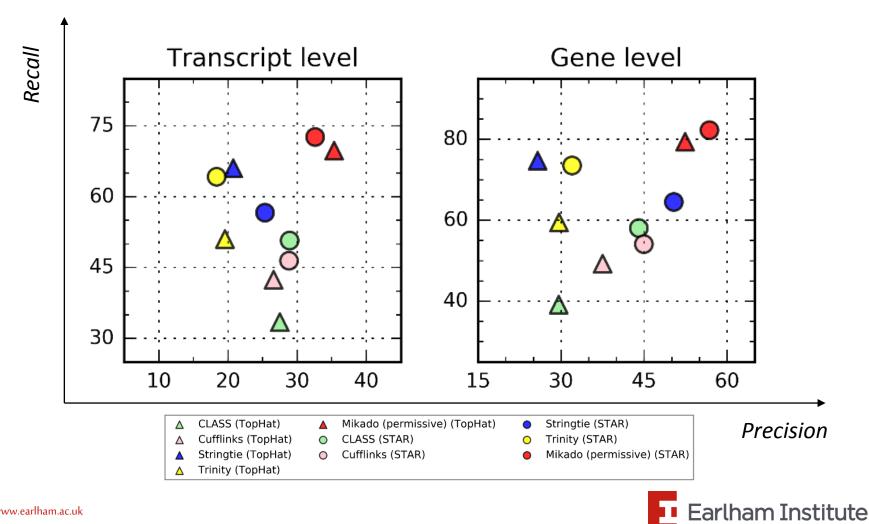
# ... or maximum sensitivity?

	C. elegans		A. thaliana		D. melanogaster		H. Sapiens		All species	
	z-score	Rank	z-score	Rank	z-score	Rank	z-score	Rank	z-score	Rank
Method										
StringTie (Tophat)	6.91	1	6.91	1	7.20	1	3.87	1	24.90	1
StringTie (STAR)	3.85	4	3.85	3	5.78	2	3.04	3	16.52	2
Trinity (STAR)	-5.22	7	5.83	2	-0.43		0.97	6	1.14	3
CLASS (STAR)	4.16	2	1.82	4	-8.74	<i>4000000</i>	3.59	2	0.84	4
CLASS (Tophat)	3.89	3	-11.33	411111111	3.24	3	2.33	4	-1.87	5
Cufflinks (STAR)	-4.05		-1.28	5	-3.32	6	1.54	5	-7.10	6
Cufflinks (Tophat)	-5.34	mmm	-4.24	7	-3.69	7	-0.52	7	-13.79	
Trinity (Tophat)	-4.20		-1.57	6	-0.04	4	-14.83	4111111116	-20.64	





#### Increased precision after filtering with Mikado





#### Increased precision after filtering with Mikado

